

KNOCK-OUT MICE: POSSIBLE SOLUTIONS TO THE GENETIC BACKGROUND AND FLANKING GENE PROBLEM 472.3



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Knockout effects in mice may be faked by incapacitating alleles flanking the targeted locus

It has been noted (R. Gerlai, *TINS* 19: 177-180, 1996) that mice carrying targeted null ("knock-out", KO) mutations differ from control animals not only by the invalidated gene, but also in the origin of any genes flanking that gene. Specifically, mutant mice derived from embryonic stem (ES) cells of the 129 strains will carry flanking alleles from that same strain. Since the 129 strains possess many alleles impairing performance in a variety of learning tasks, such flanking alleles may, with bad luck, induce spurious behavioral and neurophysiological impairments falsely attributed to the targeted mutation.

Based on the recommendations of the Banbury Conference (Neuron 19:755-759, 1997) we show here that this problem can be controlled using proper breeding schemes classically used in behavioral genetics. These breeding schemes are in any case necessary to maintain the mutation for further investigation.

As the large majority of KO mutants is based on 129 derived ES cells and phenotypes have been analyzed usually on a mixed 129 x C57BL/6 background, we use these two strains as an example. The principles remain the same for other strains.

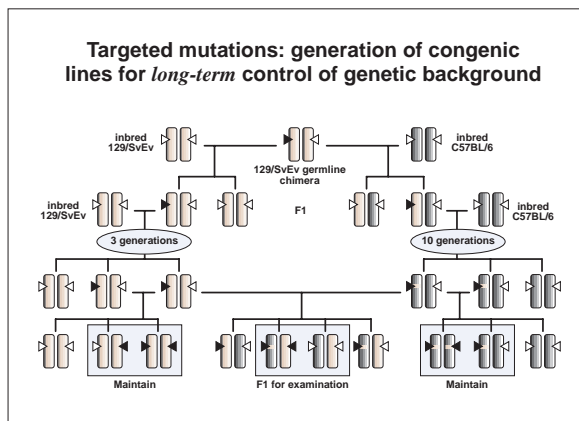
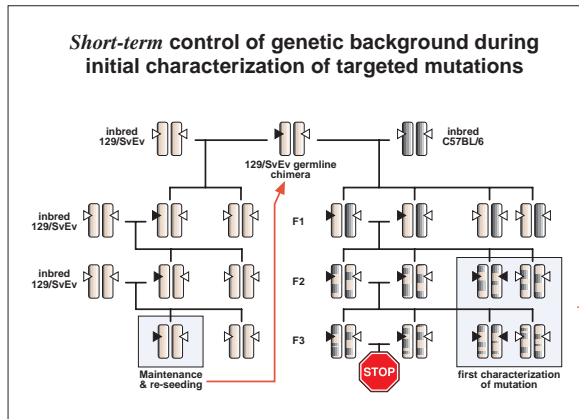


Recommendations

1. Use **proper breeding schedules**, cull litter sizes, use littermates whenever possible
2. **Measure your phenotype(s) as soon as possible in the parental strains!!!**
3. **Check your F1 generation** (obtained by mating 129 germline chimeras with C57BL/6 mice) and **compare it with the parental strains**. This will show presence or absence of hybrid vigor. If present (F1 better than parental strains), the flanking gene problem is greatly diminished (we explain that). This comparison will also detect **dominant** flanking allele and mutation effects. If wildtypes show 129 features, there may be a problem!
4. Test initially for (typically) **recessive KO-effects using an F2 on a mixed 129 x C57BL/6 background**. It is not ideal but will work in most cases. Initiate backcrossing to the parental strains as soon as possible and continue backcrossing during the analysis of phenotypes.
5. Use then one of the testing methods indicated above. The most economical (but not the fastest) solution is to obtain littermates which are **different for the targeted locus** but always **homozygous for the flanking 129 derived region**.
6. Now you may safely publish (or withdraw/correct) your data...



This needs to be done in any case



Some minor caveats for the interested reader

1. Theoretically, **F1 hybrids** are optimal for phenotypical analysis. Slight drawback: they often **mask weak phenotypes**.
2. In our experience (about 2000 mice, about 30 KO lines), a mixed background with enough 129 alleles is helpful in discovering KO phenotypes.
3. Our models apply only if there is **no interaction between targeted locus and confounding loci**
4. Be wary that a **KO dependent phenotype** is not a function of the targeted locus but a **function of the entire genome minus the targeted locus...**



Three techniques to check for flanking gene effects

